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International Union of Pharmacology. LII. Nomenclature and Molecular Relationships of Calcium-Activated Potassium Channels

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Introduction

The second major group of six/seven transmembrane potassium-selective channels consists of the K_{Ca} channels (for reviews, see Lingle, 2002; Magleby, 2003; Moczydlowski, 2004; Stocker, 2004; Cox, 2005), and Table 1 shows the International Union of Pharmacology (IUPHAR¹) names of the members of this group together with their HUGO Gene Nomenclature Committee (HGNC) designations and other commonly used names. The phylogenetic trees in Fig. 1 illustrate the fact that these channels form two well defined but only distantly related groups.

One of these groups (Fig. 1A) includes the three "small-conductance" $K_{\rm Ca}$ channels (K_{\rm Ca}2.1, 2.2, and 2.3) (Kohler et al., 1996) and the "intermediate-conductance" channel K_{Ca}3.1 (Ishii et al., 1997; Joiner et al., 1997). These channels are voltage-insensitive and are activated by low concentrations of internal Ca²⁺ (<1.0 μ M), in contrast to K_{Ca}1.1 (KCNMA1, Slo1), which is activated by both voltage and internal Ca²⁺. The three small-conductance K_{Ca} channels are sensitive to block by apamin (100 pM-10 nM), which distinguishes them from all other K_{Ca} channels. Both small- and intermediate-conductance K_{Ca} channels play important roles in many processes involving Ca²⁺-dependent signaling in both electrically excitable and nonexcitable cells. They do not bind Ca²⁺ directly but rather detect Ca²⁺ by virtue of calmodulin, which is constitutively bound to the C-terminal region (Xia et al., 1998; Fanger et al., 1999). Binding

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¹ Abbreviations: IUPHAR, International Union of Pharmacology; HGNC, HUGO Gene Nomenclature Committee; RCK, regulator of K⁺ conductance. of calcium to this calmodulin results in conformational changes that are in turn responsible for channel gating (Schumacher et al., 2001).

The tree shown in Fig. 1B illustrates the sequence relationships within the second group of K_{Ca} channels, which includes K_{Ca}1.1 (Slo or Slo1), K_{Ca}4.1 (Slack or Slo2.2), $K_{Ca}4.2$ (Slick or Slo2.1), and $K_{Ca}5.1$ (Slo3). K_{Ca} 1.1 has been extensively studied in the brain, cochlea, and muscle, and alternate splicing of its mRNA is known to produce considerable functional diversity (Weiger et al., 2002; Faber and Sah, 2003). Unlike the K_{Ca}^2 and K_{Ca}^3 channels, binding of calcium by K_{Ca}^1 .1 is not dependent on its association with calmodulin but is thought to be mediated by at least three divalent cation binding sites in the cytoplasmic carboxyl domain of each channel subunit. Two independent high-affinity Ca²⁺ binding sites are formed by a negatively charged segment in the distal carboxyl terminal portion, termed the "calcium bowl" (Schreiber and Salkoff, 1997) and within the first RCK domain encoded by the proximal C-terminal portion (Bao et al., 2002; Xia et al., 2002). A third low-affinity divalent cation binding site is also found in the first RCK domain (Shi et al., 2002), which contributes to activation by Mg^{2+} and Ca^{2+} at high concentrations (>1 mM).

The three other members of this group, $K_{Ca}4.1$, 4.2, and 5.2 (Joiner et al., 1997; Schreiber et al., 1998; Yuan et al., 2003), were all included in the K_{Ca} nomenclature since they all are clearly members of this structurally related group of genes. However, much more is now known about the functional properties of the members of this gene family than was known when these names were assigned several years ago, and this presents a possible conundrum for a nomenclature based on functional rather than structural similarity. Unlike the founding member K_{Ca} 1.1, which is in fact activated by internal Ca^{2+} , none of the other members of this group seems to be similarly Ca^{2+} activated. In fact, for the most part, these three are insensitive to internal Ca²⁺. $K_{Ca}4.2$ and $K_{Ca}4.1$ are activated by internal Na⁺ and Cl⁻ (Yuan et al., 2003),

A.D.W. and H.W. serve as the Subcommittee on K_{Ca} Channels of the Nomenclature Committee of the International Union of Pharmacology.

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TABLE 1 K_{Ca} channels IUPHAR names of the members of the K_{Ca} group of potassium channels are shown, together with their HGNC designations and other commonly used names.

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HGNC	Other
KCNMA1	Slo, Slo1, BK
KCNN1	$SK_{Ca}1$
KCNN2	SK_{Ca}^{-2}
KCNN3	SK _{Ca} 3
KCNN4	IK _{Ca} 1
KCNT1	Slack, Slo2.2
KCNT2	Slick, Slo2.1
KCNU1	Slo3
	HGNC KCNMA1 KCNN1 KCNN2 KCNN3 KCNN4 KCNT1 KCNT2

BK, big-conductance K^+ channel; SK, small-conductance K^+ channel; IK, intermediate-conductance K^+ channel.

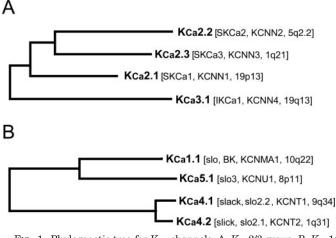


FIG. 1. Phylogenetic tree for $\rm K_{Ca}$ channels. A, $\rm K_{Ca}2/3$ group. B, $\rm K_{Ca}1/4/5$ group. Amino acid sequence alignments and phylogenetic analysis for these two groups of four human $\rm K_{Ca}$ channels were generated as described in the legend for Fig. 1 of "International of Union of Pharmacology LIII. Nomenclature and Molecular Relationships of Voltage-Gated Potassium Channels." No new channels have been added to these topologies since they appeared in the earlier edition of this compendium. IUPHAR and HGNC names of the genes are shown together with other commonly used names and their chromosomal localization.

and $K_{Ca}5.1$ is activated by internal alkalization (OH⁻) (Schreiber et al., 1998). Therefore, although they are structurally related to $K_{Ca}1.1$, these three channels cannot correctly be described as "calcium-activated" channels based on functional criteria. This may be a subject for discussion among researchers in this field and those bodies responsible for standardizing gene nomenclature.

Tables 2 through 9 present the K_{Ca} 1.1 through K_{Ca} 5.1 channels.

REFERENCES

- Bao L, Rapin AM, Holmstrand EC, and Cox DH (2002) Elimination of the BK_{Ca} channel's high-affinity Ca²⁺ sensitivity. J Gen Physiol **120:**173–189.
- Cox DH (2005) The BK_{Ca} channel's Ca²⁺-binding sites, multiple sites, multiple ions. J Gen Physiol 125:253–255.
- Faber ESL and Sah P (2003) Calcium-activated potassium channels: multiple contributions to neuronal function. *Neuroscientist* **9:**181–194.
- Fanger CM, Ghanshani S, Logsdon NJ, Rauer H, Kalman K, Zhou J, Beckingham K, Chandy KG, Cahalan MD, and Aiyar J (1999) Calmodulin mediates calcium-dependent activation of the intermediate conductance K_{Ca} channel, IK_{Ca} 1. J Biol Chem **274**:5746–5754.
- Ishii TM, Silvia C, Hirschberg B, Bond CT, Adelman JP, and Maylie J (1997) A human intermediate conductance calcium-activated potassium channel. Proc Natl Acad Sci USA 94:11651–11656.
- Joiner WJ, Tang MD, Wang LY, Dworetzky SI, Boissard CG, Gan L, Gribkoff VK, and Kaczmarek LK (1998) Formation of intermediate-conductance calciumactivated potassium channels by interaction of Slack and Slo subunits. *Nat Neu*rosci 1:462–469.
- Joiner WJ, Wang LY, Tang MD, and Kaczmarek LK (1997) hSK4, a member of a novel subfamily of calcium-activated potassium channels. *Proc Natl Acad Sci USA* 94:11013–11018.
- Kohler M, Hirschberg B, Bond CT, Kinzie JM, Marrion NV, Maylie J, and Adelman JP (1996) Small-conductance, calcium-activated potassium channels from mammalian brain. Science 273:1709-1714.
- Lingle CJ (2002) Setting the stage for molecular dissection of the regulatory components of BK channels. J Gen Physiol **120:**261–265.
- Magleby KL (2003) Gating mechanism of BK (Slo1) channels: so near, yet so far. J Gen Physiol 121:81–96. Moczydlowski EG (2004) BK channel news: full coverage on the calcium bowl. J Gen
- *Physiol* **123:**471–473. Schreiber M and Salkoff L (1997) A novel calcium-sensing domain in the BK channel.
- Biophys J 73:1355–1363.
 Schreiber M, Wei A, Yuan A, Gaut J, Saito M, and Salkoff L (1998) Slo3, a novel pH-sensitive K⁺ channel from mammalian spermatocytes. J Biol Chem 273:3509–3516.
- Schumacher MA, Rivard AF, Bachinger HP, and Adelman JP (2001) Structure of the gating domain of a Ca²⁺-activated K⁺ channel complexed with Ca²⁺/calmodulin. *Nature (Lond)* **410:**1120–1124.
- Shi J, Krishnamoorthy G, Yang Y, Hu L, Chaturvedi N, Harilal D, Qin J, and Cui J (2002) Mechanism of magnesium activation of calcium-activated potassium channels. *Nature (Lond)* 418:876-880.
- nels. Nature (Lond) **418**:876–880. Stocker M (2004) Ca²⁺-activated K+ channels: molecular determinants and function of the SK family. Nat Rev Neurosci **5**:758–770.
- Weiger TM, Hermann A, and Levitan IB (2002) Modulation of calcium-activated potassium channels. J Comp Physiol A Neuroethol Sens Neural Behav Physiol 188:79-87.
- Xia XM, Fakler B, Rivard A, Wayman G, Johnson-Pais T, Keen JE, Ishii T, Hirschberg B, Bond CT, Lutsenko S, et al. (1998) Mechanism of calcium gating in small-conductance calcium-activated potassium channels. *Nature (Lond)* 395:503– 507.
- Xia XM, Zeng X, and Lingle CJ (2002) Multiple regulatory sites in large-conductance calcium-activated potassium channels. *Nature (Lond)* **418**:880–884.
- Yuan A, Santi CM, Wei A, Wang ZW, Pollak K, Nonet M, Kaczmarek L, Crowder CM, and Salkoff L (2003) The sodium-activated potassium channel is encoded by a member of the Slo gene family. *Neuron* 37:765–773.



NOMENCLATURE AND MOLECULAR RELATIONSHIPS OF K_{ca} CHANNELS TABLE 2

 $K_{Ca}1.1$ channels

conductance calcium and voltage activated potassium channel

Channel	name
Descripti	ion
Other na	ames
Molecula	r information

Description	Large conductance, calcium- and voltage-activated potassium channel
Other names	Slo ^{1–8} , Slo1, BK channel, maxi K ⁺ channel
Molecular information	Human: 1182aa, NM_001014797 (transcript variant 1), chr. 10q22.3, ⁶ KCNMA1
	Mouse: 1171aa, NM_010610, chr. 14;
	Rat: 1243aa, NM_031828, chr. 15p16
Associated subunits	KCNMB1-4, ²⁹ BK- β , ^{9,10} heteromeric association with Slack (rat), ¹¹ β 2-adrenergic receptor ²³
Functional assays	Voltage clamp, membrane potential, radioligand binding
Current	Maxi K^+ calcium-activated current in cochlea, smooth muscle, neurones in brain
Conductance	260pS^{2-8}
Ion selectivity	$P_{\rm K}/P_{\rm Na} > 50$
Activation	Calcium and voltage
Inactivation	Inactivating K_{Ca} 1.1 channels have been studied extensively in chromaffin cells and have been reported in other cell types ^{30,31} ; inactivation is conferred by the β 2- and β 3-subunits
Activators	Intracellular calcium, NS1608 and NS1619, ¹² BMS204352, ¹³ DHS-1, ¹⁴ estradiol, ¹⁶ Mg ²⁺ (1–10 nM) ²⁷
Gating inhibitors	None
Blockers	TEA (0.14 mM), charybdotoxin (2.9 nM), and iberiotoxin (1.7 nM) ¹⁷ ; paxilline (1.9 nM) ¹⁵ ; slotoxin (1.5 nM) ¹⁸ ; BmP09 Chinese scorpion toxin (27 nM) ²⁸
Radioligands	$[^{125}I]$ charybdotoxin ($K_{\rm d}=34~{\rm pM}), {}^{19}[^{125}I]$ iberiotoxin-D17Y/Y36F mutant ($K_{\rm d}=5~{\rm pM}), {}^{20}[^{19}F]$ racemic BMS204352 ¹³
Channel distribution	Ubiquitous, brain (cerebellum, habenula, striatum, olfactory bulb, neocortex, granule and pyramidal cells of the hippocampus), skeletal muscle, smooth muscle (vascular, uterine, gastric, bladder), adrenal cortex, cochlear hair cells, odontoblasts, pancreatic islet cells, colonic and kidney epithelium
Physiological functions	Pleiotropic, selectivity coupled with N-type, voltage-activated calcium channels to mediate fast afterhyperpolarization in neurones, electrical tuning of nonspiking properties of cochlear hair cells, presynaptic regulation of neurotransmitter release, effector of calcium sparks in smooth muscles
Mutations and pathophysiology	Mouse knockouts of α - and β -subunits viable, ataxia, ²⁶ defects in audition, ²⁵ incontinence, ^{24,32} erectile dysfunction ³³
Pharmacological significance	Channel openers may have applications in stroke, epilepsy, bladder over-reactivity, asthma, hypertension, gastric hypermotility and psychoses ^{13,17,21}
Comments	Multiple alternative splice forms exist; stress hormones control alternative splicing ²²
	A, tetraethylammonium; NS1608, N-(3-(trifluoromethyl)phenyl)-N'-(2-hydroxy-5-chlorophenyl)urea; NS1619, 1-(2-hydroxy-5-trifluoro- ro-benzimidazol-2-one; BMS204352, (+/-)-(5-chloro-2-methoxyphenyl)-1,3-dihydro-3-fluoro-6(trifluoromethyl)-2H-indol-2-one.

1. Atkinson NS, Robertson GA, and Ganetzky B (1991) A component of calcium-activated potassium channels encoded by the Drosophila slo locus. Science 253:551-555. 2. Butler A, Tsunoda S, McCobb DP, Wei A, and Salkoff L (1993) mSlo, a complex mouse gene encoding 'maxi' calcium-activated potassium channels. Science 261:221-224. 3. Knaus HG, Garcia-Calvo M, Kaczorowski GJ, and Garcia, ML (1994) Subunit composition of the high conductance calcium-activated potassium channel from smooth

muscle, a representative of the mSlo and slowpoke family of potassium channels. J Biol Chem 269:3921-3924.

 $K_{Ca}1.1$

4. Pallanck L and Ganetzky B (1994) Cloning and characterization of human and mouse homologs of the Drosophila calcium-activated potassium channel gene, slowpoke. Hum Mol Genet 3: 1239-1243.

5. Lagrutta A, Shen KZ, North RA, and Adelman JP (1994) Functional differences among alternatively spliced variants of Slowpoke, a Drosophila calcium-activated potassium channel. J Biol Chem 269:20347-20351.

6. Tseng-Crank J, Foster CD, Krause JD, Mertz R, Godinot N, DiChiara TJ, and Reinhart PH (1994) Cloning, expression, and distribution of functionally distinct Ca (2+)-activated K+ channel isoforms from human brain. Neuron 13:1315-1330.

7. Wei A, Solaro C, Lingle C, and Salkoff L (1994) Calcium sensitivity of BK-type KCa channels determined by a separable domain. Neuron 13:671-681.

8. Meera P, Wallner M, Song M, and Toro L (1997) Large conductance voltage- and calcium-dependent K⁺ channel, a distinct member of voltage-dependent ion channels with seven N-terminal transmembrane segments (S0-S6), an extracellular N terminus, and an intracellular (S9-S10) C terminus. *Proc Natl Acad Sci USA* **94**:14066–14071. Jiang Z, Wallner M, Meera P, and Toro L (1999) Human and rodent MaxiK channel beta-subunit genes: cloning and characterization. *Genomics* 55:57–67.
 Weiger TM, Holmqvist MH, Levitan IB, Clark FT, Sprague S, Huang WJ, Ge P, Wang C, Lawson D, Jurman ME, et al. (2000) A novel nervous system beta subunit that downregulates human large conductance calcium-dependent potassium channels. *J Neurosci* 20:3563–3570.
 Joiner WJ, Tang MD, Wang LY, Dworetzky SI, Boissard CG, Gan L, Gribkoff VK, and Kaczmarek LK (1998) Formation of intermediate-conductance calcium-activated

potassium channels by interaction of Slack and Slo subunits Nat Neurosci 1:462-469.

12. Strobaek D, Christophersen P, Holm NR, Moldt P, Ahring PK, Johansen TE, and Olesen SP (1996) Modulation of the Ca²⁺-dependent K⁺ channel, hslo, by the substituted diphenylurea NS 1608, paxilline and internal Ca²⁺. Neuropharmacology 35:903-914.

13. Gribkoff VK, Starrett JE, Dworetzky SI, Hewawasam P, Boissard CG, Cook DA, Frantz SW, Heman K, Hibbard JR, Huston K, et al. (2001) Targeting acute ischemic stroke with a calcium-sensitive opener of maxi-Kpotassium channels. Nat Med 7:471-477. 14. McManus OB, Harris GH, Giangiacomo KM, Feigenbaum P, Reuben JP, Addy ME, Burka JF, Kaczorowski GJ, and Garcia, ML (1993) An activator of calcium-

dependent potassium channels isolated from a medicinal herb. Biochemistry 32:6128-6133. 15. Sanchez M and McManus OB (1996) Paxilline inhibition of the alpha-subunit of the high conductance calcium-activated potassium channel. Neuropharmacology

35:963–968.

16. Valverde MA, Rojas P, Amigo J, Cosmelli D, Orio P, Bahamonde MI, Mann GE, Vergara C and Latorre R (1999) Acute activation of Maxi-K channels (hSlo) by estradiol binding to the beta subunit. Science 285:1929-1931.

17. Kaczorowski GJ, Knaus HG, Leonard RJ, McManus OB, and Garcia ML (1996) High-conductance calcium-activated potassium channels; structure, pharmacology, and function. J Bioenerg Biomembr 28:255-267.

18. Garcia-Valdes J, Zamudio FZ, Toro L, and Possani LD (2001) Slotoxin, alphaKTx1.11, a new scorpion peptide blocker of MaxiK channels that differentiates between alpha and alpha + beta (beta1 or beta4) complexes. FEBS Lett 505:369-373. 19. Garcia-Calvo M, Knaus HG, McManus OB, Giangiacomo KM, Kaczorowski GJ, and Garcia ML (1994) Purification and reconstitution of the high-conductance,

calcium-activated potassium channel from tracheal smooth muscle. J Biol Chem 269:676-682. 20. Koschak A, Koch RO, Liu J, Kaczorowski GJ, Reinhart PH, Garcia ML, and Knaus HG (1997) [¹²⁵I]Iberiotoxin-D19Y/Y36F, the first selective, high specific activity

radioligand for high-conductance calcium-activated potassium channels. Biochemistry 36:1943-1952. 21. Coghlan MJ, Carrol WA, and Gopalakrishnan M (2001) Recent developments in the biology and medicinal chemistry of potassium channel modulators: update from

a decade of progress. J Med Chem 44:1-27. 22. Xie J and McCobb DP (1998) Control of alternative splicing of potassium channels by stress hormones. Science 280:443-446.

23. Liu G, Shi J, Yang L, Cao L, Park SM, Cui J, and Marx SO (2004) Assembly of a Ca2+-dependent BK channel signaling complex by binding to beta2 adrenergic receptor. EMBO J 23:2196-2205.

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24. Meredith AL, Thorneloe KS, Werner ME, Nelson MT, and Aldrich RW (2004) Overactive bladder and incontinence in the absence of the BK large conductance Ca2+-activated K + channel. J Biol Chem 279:36746-36752.

25. Ruttiger L, Sausbier M, Zimmermann U, Winter H, Braig C, Engel J, Knirsch M, Arntz C, Langer P, et al. (2004) Deletion of the Ca2+-activated potassium (BK) alpha-subunit but not the BK beta1-subunit leads to progressive hearing loss. Proc Natl Acad Sci USA 101:12922-12927.

26. Sausbier M, Hu H, Arntz C, Feil S, Kamm S, Adelsberger H, Sausbier U, Sailer CA, Feil R, Hofmann F, et al. (2004) Cerebellar ataxia and Purkinje cell dysfunction caused by Ca2+-activated K+ channel deficiency. Proc Natl Acad Sci USA 101:9474-9478.

27. Shi J, Krishnamoorthy G, Yang Y, Hu L, Chaturvedi N, Harilal D, Qin J, and Cui J (2002) Mechanism of magnesium activation of calcium-activated potassium channels. Nature (Lond) 418:876-880.

28. Yao J, Chen X, Li H, Zhou Y, Yao L, Wu G, Zhang N, Zhou Z, Xu T, Wu H, et al. (2005) BmP09, a long-chain scorpion peptide blocker of BK channels, J Biol Chem 280:14819-14828.

29. Brenner R, Jegla TJ, Wickenden A, Liu Y, and Aldrich RW (2000) Cloning and functional characterization of novel BK potassium channel beta subunits, hKCNMB3 and hKCNMB4. J Biol Chem 275:6453-6461.

30. Xia XM, Ding JP, and Lingle CJ (1999) Molecular basis for the inactivation of Ca2+ and voltage-dependent BK channels in adrenal chromaffin cells and rat insulinoma tumor cells. J Neurosci 19:5255–5264.

31. Lingle CJ, Solaro CR, Prakriya M, and Ding JP (1996) Calcium-activated potassium channels in adrenal chromaffin cells. Ion Channels 4:261–301. 32. Petkov GV, Bonev AD, Heppner TJ, Brenner R, Aldrich RW, and Nelson MT (2001) Related beta1-subunit of the Ca2+-activated K+ channel regulates contractile activity of mouse urinary bladder smooth muscle. J Physiol 537:443-452.

33. Werner ME, Zvara P, Meredith AL, Aldrich RW, and Nelson MT (2005) Erectile dysfunction in mice lacking the large conductance calcium-activated potassium (BK) channel. J Physiol 567 (Pt 2):545-556.

TABLE 3 K_{α} 2.1 channels

	$K_{Ca}2.1$ channels
Channel name	K _{Ca} 2.1
Description	Small-conductance, calcium-activated potassium channel; activated via a calmodulin-dependent mechanism
Other names	SK1 ^{1,2} , SKCa1
Molecular information	Human: 543aa, NM_002248, chr. 19p13.1, ³ KCNN1
	Mouse: 580aa, NM_032397, chr. 8
	Rat: 536aa, NM_019313, chr. 16p14
Associated subunits	Calmodulin tightly complexed to C terminus ⁴
Functional assays	Electrophysiology
Current	Small-conductance, calcium-activated K ⁺ current in neurones ¹
Conductance	9.2pS (symmetric K ⁺), 2–3pS (normal Ringer)
Ion selectivity	K ⁺ -selective
Activation	Activated by intracellular Ca ²⁺ ($K_{\rm d} = 0.7 \ \mu {\rm M}, n_{\rm H} = 4$) ⁴
Inactivation	None
Activators	Ca^{2+} , EBIO (630 μ M), ⁵ NS309 (30 nM), ⁶ riluzole (2 μ M)
Gating inhibitors	None
Blockers	UCL1684 (1 nM), ⁷ apamin (8 nM), ⁸ tamapin (42 nM), ⁹ leiurotoxin/scyllatoxin (325 nM), ¹⁰ dequalinium (400 nM), leiurotoxin-Dab7 (6 μ M), ¹⁰ fluoxetine (7 μ M), tubocurarine (23 μ m), biciculline (1.1 μ M) ¹⁴
Radioligands	[¹²⁵ I]apamin ¹¹
Channel distribution	Brain amygdala > hippocampus, caudate nucleus, foetal brain > cerebellum > thalamus, substantia nigra, spinal cord, pituitary gland), oligodendroglioma, glioblastoma, gastric tumour, aorta ^{4,12}
Physiological functions	Involved in the afterhyperpolarization in vertebrate neurones
Mutations and pathophysiology	Not established
Pharmacological significance	Modulators of SK channel subtypes may have potential use in the treatment of myotonic muscular dystrophy, gastrointestinal dysmotility, memory disorders, epilepsy narcolepsy, and alcohol intoxication ¹³
Comments	Channel is voltage-independent and weakly rectifying; intron-exon structure of $K_{\rm Ca}2.1-K_{\rm Ca}2.3~(\rm SK)$ and $K_{\rm Ca}3.1~(\rm IK)$ genes are conserved

aa, amino acids; chr., chromosome; NS309, 6,7-dichloro-1H-indole-2,3-dione-3-oxime; SK, small-conductance K⁺ channel; IK, intermediate-conductance K⁺ channel; EBIO, 1-ethyl-2-benzimidazolinone; UCL1684, 6,12,19,20,25,26-hexahycro-5,27:13,18:21,24-trietheno-11,7-methano-7H-dibenzo[b,n] [1,5,12,16] tetraazacyclotricosine-5,13dilum ditrifluoroacetate

1. Kohler M, Hirschberg B, Bond CT, Kinzie JM, Marrion NV, Maylie J, and Adelman JP (1996) Small-conductance, calcium-activated potassium channels from mammalian brain. Science 273:1709-1714.

2. Litt M, LaMorticella D, Bond CT, and Adelman JP (1999) Gene structure and chromosome mapping of the human small-conductance calcium-activated potassium channel SK1gene (KCNN1). Cytogenet Cell Genet 86:70–73.

3. Ghanshani S, Wulff H, Miller MJ, Rohm H, Neben A, Gutman GA, Cahalan MD, and Chandy KG (2000) Up-regulation of the IKCa1 potassium channel during T-cell activation. Molecular mechanism and functional consequences. J Biol Chem 275:37137-37149.

4. Xia XM, Fakler B, Rivard A, Wayman G, Johnson-Pais T, Keen JE, Ishii T, Hirschberg B, Bond CT, Lutsenko S, et al. (1998) Mechanism of calcium gating in small-conductance calcium-activated potassium channels. Nature (Lond) 395:503-507.

5. Dale TJ, Cryan JE, Chen MX, Trezise DJ (2002) Partial apamin sensitivity of human small conductance Ca²⁺-activated K⁺ channels stable expressed in Chinese hamster ovary cells. Naunyn Schmiedeberg's Arch Pharmacol 366:470-477.
6. Strobaek D, Teuber L, Jorgensen TD, Ahring PK, Kaer K, Hansen RS, Olesen SP, Christophersen P, and Skaaning-Jensen B (2004) Activation of human IK and SK

 Ca^{2} -activated K⁺ channels by NS309 (6,7-dichloro-1H-indole-2,3-dione3-oxime). Biochim Biophys Acta 1665:1-5.

7. Strobaek D, Jorgensen TD, Christophersen P, Ahring PK, and Olesen S-P (2000) Pharmacological characterization of small-conductance Ca²⁺-activated K⁺ channels stably expressed in HEK 293 cells. Br J Pharmacol 129:991-999.

8. Shah M and Haylett DG (2000) The Pharmacology of hSK1 Ca²⁺ activated K⁺ channels expressed in mammalian cell lines. Br J Pharmacol 129:627–630.

9. Pedarzani P, D'hoedt D, Doorty KB, Wadsworth JD, Joseph JS, Jeyaseelan K, Kini RM, Gadre SV, Sapatnekar SM, Stocker M, et al. (2002) Tamapin, a venompeptide from the Indian red scorpion (Mesobuthus tamulus) that targets small conductance Ca²⁺-activated K⁺ channels and after hyperpolarization currents in central neurons. J Biol Chem 277:46101-46109.

10. Shakkottai VG, Regaya I, Wulff H, Fajloun Z, Tomita H, Fathallah M, Cahalan MD, Gargus JJ, Sabatier JM, and Chandy KG (2001) Design and characterization of a highly selective peptide inhibitor of the small conductance calcium-activated K⁺ channel, SKCa2. J Biol Chem 276:43145-43151.

11. Romey G, Hugues M, Schmid-Antomarchi H, and Lazdunski M (1984) Apamin: a specific toxin to study a class of Ca²⁺-dependent K⁺ channels. J Physiol (Paris) 79:259-264. 12. Stocker M and Pedarzani P (2000) Differential distribution of three Ca²⁺-activated K⁺ channel subunits, SK1, SK2, and SK3, in the adult rat central nervous system. Mol Cell Neurosci 15:476-493.

13. Coghlan MJ, Carrol WA, and Gopalakrishnan M (2001) Recent developments in the biology and medicinal chemistry of potassium channel modulators: update from a decade of progress. J Med Chem 44:1-27.

14. Khawaled R, Bruening-Wright A, Adelman JP, and Maylie J (1999) Bicuculline block of small-conductance calcium-activated potassium channels. Pflugers Arch Eur J Physiol 438:314-321.



NOMENCLATURE AND MOLECULAR RELATIONSHIPS OF K_{ca} CHANNELS TABLE 4

 $K_{Ca}2.2$ channels

Human: 579aa, NM_021614 (transcript variant 1), chr. 5q22.3,² KCNN2

EBIO,⁶ chlorzoxazone, zoxazolamine,⁷ NS309 (30 nM),⁸ riluzole (2 μ M)

Underlies the medium afterhyperpolarization in vertebrate neurones^{18,19}

Small-conductance, calcium-activated potassium channel; activated via a calmodulin-dependent mechanism

Calmodulin tightly complexed to C terminus,^{3,4} protein kinase CK2 and protein phosphatase 2A²³

Small-conductance, calcium-activated K^+ current in neurones possibly underlies the medium I_{AHP}

Tamapin (24 pM),⁹ apamin (60–200 pM),^{1,10} leiurotoxin/scyllatoxin (200 pM), leiuritoxin-Dab7 (3.8 nM), PO5 (22 nM), Tskappa (80 nM), Pi1-OH (>1 μ M), Pi1-NH2 (100 nM), and maurotoxin (1 μ M),¹¹ UCL1684 (250pM),¹² tubocurarine (5 μ M)¹⁰; with micromolar affinity: amitriptyline, carbamazepine, chlorpromazine, cyproheptadine, fluoxetine, imipramine, tacrine, trifluperazine,¹³

Brain (spinal cord > hippocampus, cerebellum > amygdala > foetal brain > corpus callosum, thalamus, caudate nucleus, substantia nigra),¹⁵ pituitary gland, melanocyte, melanoma, germ cell tumor, prostate, oligodendroglioma, lung, Jurkat T cells,¹⁶ liver, heart,¹⁷ skeletal muscle,

Char	nnel name
Desc	ription
Othe	er names
Mole	cular information
Func	ciated subunits tional assays
Curr	ent

Conductance Ion selectivity Activation Inactivation Activators Gating inhibitors Blockers

Radioligands Channel distribution

Mutations and pathophysiology

Pharmacological significance

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Physiological functions

Comments

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Dominant-negative suppression of $K_{Ca}2.2$ channels in deep cerebellar nuclei in a transgenic mouse causes cerebellar ataxia²⁰ Modulators of SK channel subtypes may have potential use in the treatment of myotonic muscular dystrophy, gastrointestinal dysmotility, memory disorders, epilepsy, narcolepsy, and alcohol

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Comments

intoxication²¹; K_{Ca}2.2 openers have been proposed for the treatment of cerebellar ataxia²⁰ The channel is voltage-independent and weakly rectifying; shared intron-exon structure with members of the K_{Ca}^2 and K_{Ca}^3 subfamilies² aa, amino acids; chr., chromosome; NS309, 6,7-dichloro-1H-indole-2,3-dione-3-oxime; SK, small-conductance K⁺ channel; EBIO, 1-ethyl-2-benzimidazolinone; UCL1684,

 $K_{Ca}2.2$

SK2,¹ SKCa2

Electrophysiology

biciculline $(1.1 \ \mu M)^2$

¹²⁵I]apamin¹⁴

mvometrium

 K^+ -selective⁵

None

None

Mouse: 574aa, NM_080465, chr. 18 Rat: 580aa, NM_019314, chr. 18q11

current in hippocampal neurones 9.9pS (symmetric K⁺), 2–3pS (normal Ringer)⁵

Activated by intracellular Ca²⁺ ($K_{\rm d}$ = 0.6 μ M, $n_{\rm H}$ = 4)⁵

- 6,12,19,20,25,26-hexahycro-5,27:13,18:21,24-trietheno-11,7-methano-7H-dibenzo [b,n] [1,5,12,16] tetraazacyclotricosine-5,13-dilum ditrifluoroacetate. 1. Kohler M, Hirschberg B, Bond CT, Kinzie JM, Marrion NV, Maylie J, and Adelman JP (1996) Small-conductance, calcium-activated potassium channels from
- mammalian brain. Science 273:1709-1714. 2. Ghanshani S, Wulff H, Miller MJ, Rohm H, Neben A, Gutman GA, Cahalan MD, and Chandy KG (2000) Up-regulation of the IKCa1 potassium channel during T-cell
- activation. Molecular mechanism and functional consequences. J Biol Chem 275:37137-37149. 3. Xia XM, Fakler B, Rivard A, Wayman G, Johnson-Pais T, Keen JE, Ishii T, Hirschberg B, Bond CT, Lutsenko S, et al. (1998) Mechanism of calcium gating in
- small-conductance calcium-activated potassium channels. Nature (Lond) 395:503-507. 4. Schumacher MA, Rivard AF, Bachinger HP, and Adelman JP (2001) Structure of the gating domain of a Ca²⁺-activated K⁺ channel complexed with Ca²⁺/calmodulin. Nature (Lond) 410:1120-1124
- 5. Hirschberg B, Maylie J, Adelman JP, and Marrion NV (1998) Gating of recombinant small-conductance Ca-activated K⁺ channels by calcium. J Gen Physiol 111:565–581. 6. Pedarzani P, Mosbacher J, Rivard A, Cingolani LA, Oliver D, Stocker M, Adelman JP, and Fakler B (2001) Control of electrical activity in central neurons by modulating the gating of small-conductance Ca²⁺-activated K⁺ channels. J Biol Chem **276:**9762–9769.
- 7. Cao Y, Dreixler JC, Roizen JD, Roberts MT, and Houamed KM (2001) Modulation of recombinant small-conductance Ca²⁺-activated K⁺ channels by the muscle relaxant
- 8. Stroback D, Teuber L, Jorgensen TD, Ahring PK, Kaer K, Hansen RS, Olesen SP, Christophersen P, and Skaaning-Jensen B (2004) Activation of human IK and SK Ca²⁺ -activated K⁺ channels by NS309 (6.7-dichloro-1H-indole-2 3-dione-2 oximo). *Picekiw Picek* 4 (1997)
- ²⁺ -activated K⁺ channels by NS309 (6,7-dichloro-1H-indole-2,3-dione3-oxime). Biochim Biophys Acta 1665:1–5.
 9. Pedarzani P, D'hoedt D, Doorty KB, Wadsworth JD, Joseph JS, Jeyaseelan K, Kini RM, Gadre SV, Sapatnekar SM, Stocker M, et al. (2002) Tamapin, a venompeptide from the Indian red scorpion (Mesobuthus tamulus) that targets small conductance Ca²⁺ activated K⁺ channels and after hyperpolarization currents in central neurons. J Biol Chem 277:46101-46109.
- 10. Ishii TM, Maylie J, and Adelman JP (1997) Determinants of apamin and d-tubocurarine block in SK potassium channels. J Biol Chem 272:23195-23200.
- 11. Shakkottai VG, Regaya I, Wulff H, Fajloun Z, Tomita H, Fathallah M, Cahalan MD, Gargus JJ, Sabatier JM, and Chandy KG (2001) Design and characterization of a highly selective peptide inhibitor of the small conductance calcium-activated K⁺ channel, SKCa2. J Biol Chem 276:43145-43151.
- 12. Fanger CM, Rauer H, Neben AL, Miller MJ, Rauer H, Wulff H, Rosa JC, Ganellin CR, Chandy KG, and Cahalan MD (2000) Calcium-activated potassium channels sustain calcium signaling in T lymphocytes. Selective blockers and manipulated channel expression levels. J Biol Chem 276: 12249-12256.
- 13. Dreixler JC, Bian J, Cao Y, Roberts MT, Roizen JD, and Houamed KM (2000) Block of rat brain recombinant SK channels by tricyclic antidepressants and related compounds. Eur J Pharmacol 401:1-7.
- 14. Romey G, Hugues M, Schmid-Antomarchi H, and Lazdunski M (1984) Apamin: a specific toxin to study a class of Ca²⁺-dependent K⁺ channels. J Physiol (Paris) 79:259-264. 15. Stocker M and Pedarzani P (2000) Differential distribution of three Ca²⁺-activated K⁺ channel subunits, SK1, SK2, and SK3, in the adult rat central nervous system. Mol Cell Neurosci 15:476-493.
- 16. Jager H, Adelman JP, and Grissmer S (2000) SK2 encodes the apamin-sensitive Ca (2+)-activated K(+) channels in the human leukemic T cell line, Jurkat. FEBS Lett 469:196-202.
- 17. Xu Y, Tuteja D, Zhang Z, Xu D, Zhang Y, Rodriguez J, Nie L, Tuxson HR, Young JN, Glatter KA, et al. (2003) Molecular identification and functional roles of a Ca²⁺-activated K⁺ channel in human and mouse hearts. *J Biol Chem* **278**:49085–49094. 18. Villalobos C, Shakkottai VG, Chandy KG, Michelhaugh SK, and Andrade R (2004) SKCa channels mediate the medium but not the slow calcium-activated
- afterhyperpolarization in cortical neurons. J Neurosci 24:3537-3542. 19. Bond CT, Herson PS, Strassmaier T, Hammond R, Stackman R, Maylie J, and Adelman JP (2004) Small conductance Ca2+-activated K+ channel knock-out mice reveal
- the identity of calcium-dependent afterhyperpolarization currents. J Neurosci 24:5301-5306. 20. Shakkottai VG, Chou CH, Oddo S, Sailer CA, Knaus HG, Gutman GA, Barish ME, LaFerla FM, and Chandy KG (2004) Enhanced neuronal excitability in the absence
- of neurodegeneration induces cerebellar ataxia. J Clin Investig 113:582-590. 21. Coghlan MJ, Carrol WA, and Gopalakrishnan M (2001) Recent developments in the biology and medicinal chemistry of potassium channel modulators: update from a decade of progress. J Med Chem 44:1-27.
- 22. Khawaled R, Bruening-Wright A, Adelman JP, and Maylie J (1999) Bicuculline block of small-conductance calcium-activated potassium channels. Pflugers Arch Eur J Physiol 438:314-321.
- 23. Bildl W, Strassmaier T, Thurm H, Andersen J, Eble S, Oliver D, Knipper M, Mann M, Schulte U, Adelman JP, et al. (2004) Protein Kinase CK2 is coassembled with small conductance Ca2+-activated K+ channels and regulates channel gating. Neuron 43:847-858.

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$X_{Ca}2.3$	chai	nnels

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	K _{Ca} 2.3 channels
Channel name	K _{Ca} 2.3
Description	Small-conductance, calcium-activated potassium channel activated via a calmodulin-dependent mechanism
Other names	SK3, ¹ hKCa3, SKCa3 ²
Molecular information	Human: 736aa, NM_002249 (transcript variant 1), chr. 1q21.3, 3,4 KCNN3
	Mouse: 731aa, NM_080466, chr. 3
	Rat: 732aa, NM_019315, chr. 2q34
Associated subunits	Calmodulin tightly complexed to C terminus ^{5,6}
Functional assays	Patch-clamp
Current	Small-conductance, calcium-activated K^+ current in neurones ⁷
Conductance	Not determined
Ion selectivity	K ⁺ -selective
Activation	Activated by intracellular Ca^{2+} ($K_d = 0.6 \text{ uM}$) ⁸
Inactivation	None
Activators	EBIO, riluzole (3 μ M), ⁹ NS309 (30 nM) ¹⁰
Gating inhibitors	None
Blockers	Leiurotoxin/scyllatoxin (1.1 nM), apamin (10 nM), PO5 (25 nM), Tskappa (197 nM), Pi1-OH (330 nM), and Pi1-NH2 (250 nM), ¹¹ UCL1684 (9.5 nM) ¹² ; with micromolar affinity: bicuculline, ⁹ amitriptyline,
	fluoxetine, desipramine, imipramine, nortriptyline, fluphenazine, promethazine, chlorpromazine
Radioligands	[¹²⁵ I]apamine ¹³
Channel distribution	Brain (substantia nigra > amygdala, caudate nucleus, thalamus, hippocampus, ventral tegmental area, cerebellum, spinal cord > corpus callosum, foetal brain), lymphocytes (germinal center B cells, tonsillar B cells, Burkitt's lymphoma, microglia), skeletal muscle (increased denervated muscle, myotonic dystrophy), myometrium, prostate, kidney, heart, pituitary gland, liver, pancreas, colon, germinal cells, head, neck, ovary, vascular endothelium ^{1,3,14–19}
Physiological functions	Involved in the afterhyperpolarization in vertebrate neurones ^{7,17} (any newer comments on this?)
Mutations and pathophysiology	Longer polyglutamine repeats are over-represented in schizophrenic (especially negative-symptom form) ^{2,18} individuals and in patients with anorexia nervosa ²⁰ and spinocerebellar ataxia ²¹ ; a fourbase deletion has been found in a patient with schizophrenia ²² that truncates the protein just before the S1 segment and causes dominant-negative suppression of endogenous SK channels ²³ ; protein and mRNA levels are increased in skeletal muscle following denervation ²⁴ and in patients with myotonic muscular dystrophy ²⁵ ; involved in the endothelium-mediated vasodilation (EDHF response) ¹⁹ ; conditional knockout of K _{Ca} 2.3 leads to hypertension ²⁶ and bladder instability ²⁷
Pharmacological significance	Modulators of SK channel subtypes may have potential use in the treatment of myotonic muscular dystrophy, gastrointestinal dysmotility, memory disorders, epilepsy, narcolepsy, hypertension, ²⁶ and urinary incontinence ²⁷
Comments	Channel is voltage-independent

linone; SK, small-conductance K⁺ channel; UCL1684, 6,12,19,20,25,26-hexahycro-5,27:13,18:21,24-trietheno-11,7-methano-7H-dibenzo [b,n] [1,5,12,16] tetraazacyclotricosine-5,13-dilum ditrifluoroacetate.

1. Kohler M, Hirschberg B, Bond CT, Kinzie JM, Marrion NV, Maylie J, and Adelman JP (1996) Small-conductance, calcium-activated potassium channels from mammalian brain. Science 273:1709–1714.

2. Chandy KG, Fantino E, Wittekindt O, Kalman K, Tong LL, Ho TH, Gutman GA, Crocq MA, Ganguli R, Nimgaonkar V, et al. (1998) Isolation of a novel potassium channel gene hSKCa3 containing a polymorphic CAG repeat: a candidate for schizophrenia and bipolar disorder? Mol Psychiatry 3:32-37.

3. Dror V, Shamir E, Ghanshani S, Kimhi R, Swartz M, Barak Y, Weizman R, Avivi L, Litmanovitch T, Fantino E, et al. (1999) hKCa3/KCNN3 potassium channel gene: association of longer CAG repeats with schizophrenia in Israeli Ashkenazi Jews, expression inhuman tissues and localization to chromosome 1q21. Mol Psychiatry 4:254-260.

Sun G, Tomita H, Shakkottai VG, and Gargus JJ (2001) Genomic organization and promoter analysis of human KCNN3 gene. J Hum Genet 46:463–470.
 Xia XM, Fakler B, Rivard A, Wayman G, Johnson-Pais T, Keen JE, Ishii T, Hirschberg B, Bond CT, Lutsenko S, et al. (1998) Mechanism of calcium gating in small-conductance calcium-activated potassium channels. Nature (Lond) 395:503–507.

6. Fanger CM, Ghanshani S, Logsdon NJ, Rauer H, Kalman K, Zhou J, Beckingham K, Chandy KG, Cahalan MD, and Aiyar J (1999) Calmodulin mediates calcium-dependent activation of the intermediate conductance KCa channel, IKCa1. J Biol Chem 274:5746-5754.

7. Wolfart J, Neuhoff H, Franz O, and Roeper J (2001) Differential expression of the small-conductance, calcium-activated potassium channel SK3 is critical for pacemaker control in dopaminergic midbrain neurons. J Neurosci 21:3443-3456.

8. Barfod ET, Moore AL, and Lidofsky SD (2001) Cloning and functional expression of a liver isoform of the small conductance Ca²⁺-activated K⁺ channel SK3. Am J Physiol Cell Physiol 280:C836-C842.

9. Grunnet M, Jespersen T, Angelo K, Frokjaer-Jensen C, Klaerke DA, Olesen SP, and Jensen BS (2001) Pharmacological modulation of SK3 channels. Neuropharmacology 40:879-887.

10. Strobaek D, Teuber L, Jorgensen TD, Ahring PK, Kaer K, Hansen RS, Olesen SP, Christophersen P, and Skaaning-Jensen B (2004) Activation of human IK and SK -activated K⁺ channels by NS309 (6,7-dichloro-1H-indole-2,3-dione3-oxime). Biochim Biophys Acta 1665:1-5. 11. Shakkottai VG, Regaya I, Wulff H, Fajloun Z, Tomita H, Fathallah M, Cahalan MD, Gargus JJ, Sabatier JM, and Chandy KG (2001) Design and characterization of

a highly selective peptide inhibitor of the small conductance calcium-activated K⁺ channel, SKCa2. J Biol Chem 276:43145-43151. 12. Fanger CM, Rauer H, Neben AL, Miller MJ, Rauer H, Wulff H, Rosa JC, Ganellin CR, Chandy KG, and Cahalan MD (2000) Calcium-activated potassium channels

sustain calcium signaling in T lymphocytes. Selective blockers and manipulated channel expression levels. J Biol Chem 276: 12249–12256.
 13. Romey G, Hugues M, Schmid-Antomarchi H, and Lazdunski M (1984) Apamin: a specific toxin to study a class of Ca²⁺-dependent K⁺ channels. J Physiol (Paris) 79:259–264.
 14. Stocker M and Pedarzani P (2000) Differential distribution of three Ca²⁺-activated K⁺ channel subunits, SK1, SK2, and SK3, in the adult rat central nervous system.

Mol Cell Neurosci 15:476-493.

15. Pribnow D, Johnson-Pais T, Bond CT, Keen J, Johnson RA, Janowsky A, Silvia C, Thayer M, Maylie J, and Adelman JP (1999) Skeletal muscle and small-conductance calcium-activated potassium channels. Muscle and Nerve 22:742-750.

16. UniGene Cluster Hs0.89230; Online Mendelian Inheritance in Man (OMIM) no. 602983.

17. Hosseini R, Benton DC, Dunn PM, Jenkinson DH, and Moss GW (2001) SK3 is an important component of K (+) channels mediating the after-hyperpolarization in cultured rat SCG Neurones. J Physiol 535:323-334.

18. Cardno AG, Bowen T, Guy CA, Jones LA, McCarthy G, Williams NM, Murphy KC, Spurlock G, Gray M, Sanders RD, et al. (1999) CAG repeat length in the hKCa3 gene and symptom dimensions in schizophrenia. *Biol Psychiatry* 45:1592–1596.
 19. Burnham MP, Bychkov R, Feletou M, Richards GR, Vanhoutte PM, Weston AH, and Edwards G (2002) Characterization of an apamin-sensitive small-conductance

-activated K⁺ channel in porcine coronary artery endothelium: relevance to EDHF. Br J Pharmacol 135:1133-1143.

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20. Koronyo-Hamaoui M, Danziger Y, Frisch A, Stein D, Leor S, Laufer N, Carel C, Fennig S, Minoumi M, Apter A, et al. (2002) Association between anorexia nervosa and the hsKCa3 gene: a family-based and case control study. *Mol Psychiatry* 7:82-85.

21. Figueroa KP, Chan P, Schols L, Tanner C, Riess O, Perlman SL, Geschwind DH, and Pulst SM (2001) Association of moderate polyglutamine tract expansions in the slow calcium-activated potassium channel type 3 with ataxia. Arch Neurol 58:1649-1653.

22. Bowen T, Williams N, Norton N, Spurlock G, Wittekindt OH, Morris-Rosendahl DJ, Williams H, Brzustowicz L, Hoogendoorn B, Zammit S, Jones G, et al. (2001) Mutation screening of the KCNN3 gene reveals a rare frame shift mutation. Mol Psychiatry 6:259-260.

23. Miller MJ, Rauer H, Tomita H, Rauer H, Gargus JJ, Gutman GA, Cahalan MD, and Chandy KG (2001) Nuclear localization and dominant-negative suppression by a mutant SKCa3 N-terminal channel fragment identified in a patient with schizophrenia. J Biol Chem 276:27753-27756.

24. Neelands TR, Herson PS, Jacobson D, Adelman JP, and Maylie J (2001) Small-conductance calcium-activated potassium currents in mouse hyperexcitable denervated skeletal muscle. J Physiol 536:397-407.

25. Kimura T, Takahashi MP, Okuda Y, Kaido M, Fujimura H, Yanagihara T, and Sakoda S (2000) The expression of ion channel mRNAs in skeletal muscles from patients with myotonic muscular dystrophy. Neurosci Lett 295:93-96.

26. Taylor MS, Bonev AD, Gross TP, Eckman DM, Brayden JE, Bond CT, Adelman JP, and Nelson MT (2003) Altered expression of small-conductance Ca²⁺-activated K^+ (SK3) channels modulates arterial tone and blood pressure. Circ Res **93**:124–131. 27. Herrera GM, Pozo MJ, Zvara P, Petkov GV, Bond CT, Adelman JP, and Nelson MT (2003) Urinary bladder instability induced by selective suppression of the murine

small conductance calcium-activated potassium (SK3) channel. J Physiol 551:893-903.

TABLE 6

	$K_{Ca}3.1 \ channels$
Channel name	K _{Ca} 3.1
Description	Intermediate-conductance, calcium-activated potassium channel; activated via a calmodulin- dependent mechanism
Other names	SK41, ¹ IK1, ² Gardos channel, $K_{Ca}4$, ³ I $K_{Ca}1^4$
Molecular information	Human: 427aa, NM_002250, chr. 19q13.2, ^{4,5} KCNN4
	Mouse: 425aa, NM_008433, chr. 7
	Rat: 424aa, NM_023021, 1q21
Associated subunits	Calmodulin tightly complexed to C terminus ⁶
Functional assays	Electrophysiology
Current	Gardos channel in erythrocytes, ⁷ IK current in lymphocytes, ⁸ fibroblasts ⁹
Conductance	$11 pS^{1-3,8}$
Ion selectivity	${ m K}^+ \left(1 ight) > { m Rb}^+ \left({ m (0.96)} > { m NH}_4 ^+ \left({ m (0.17)} > { m Cs}^+ \left({ m (0.07)}^8 ight)$
Activation	Activated by intracellular Ca ²⁺ ($K_{\rm d}$ = 0.1–0.3 μ M; $n_{\rm H}$ = 1.7–4) ^{1–4,8}
Inactivation	None
Activators	EBIO, NS309 (10 nM), 10 DCEBIO (1 μ M), 11 riluzole (1 μ M), methylxanthine (theophylline, caffeine, IBMX) 12
Gating inhibitors	None
Blockers	ChTX (5 nM), ^{1-4,14,15} maurotoxin (1 nM), ¹⁵ 4-phenyl-4 <i>H</i> -pyran 11 (8 nM), ¹⁶ ICA17043 (11 nM), ¹⁷ TRAM- 34 (20 nM), ^{14,15} ChTX-Glu ^{13,32} (33 nM), ^{14,15} ShK (30 nM), ¹⁴ clotrimazole (70 nM), ^{14,15} BgK (172 nM), ¹⁴ TRAM-3 (520 nM), ¹⁵ nitredipine (900 nM), nimodipine (1 μ M), and nifedipine (4 μ M), ¹⁴ UCL1608 (4 μ M), ¹⁸ ketoconazole (30 μ M) and econazole (12 μ M), ^{14,15} cetiedil, ¹⁸ TEA (24 mM) ¹⁴
Radioligands	None
Channel distribution	Placenta, prostate, erythrocytes, ¹⁹ lymphocytes, ^{3,4} microglia, liver, foetal liver, pancreas, hematopoietic stem cells, fibroblasts, ⁹ HL60, colon, Paneth cells, ²⁰ melanomas, ²¹ proliferating smooth muscle cells, ²² vascular endothelium, ²³ lung and colonic endothelium
Physiological functions	K _{Ca} 3.1 is involved in volume regulation in erythrocytes ^{19,24} ; its expression is up-regulated during activation of lymphocytes, and specific blockers suppress lymphocyte ^{4,8,25,26} and vascular smooth muscle cell proliferation ²² ; K _{Ca} 3.1 is involved in EDHF-mediated vasodilatation ²³ and in angiogenesis ^{27,28}
Mutations and pathophysiology	T lymphocytes and erythrocytes from $K_{Ca}3.1$ knockout mouse show sever defect in volume regulation ²⁹
Pharmacological significance	K _{Ca} 3.1 blocker ICA17043 is in clinical trials for sickle cell anemia ²⁴ ; K _{Ca} 3.1 blockers are of potential use for the treatment of diarrhea ³⁰ and as immunosuppressants ^{14,31} ; TRAM-34 has been shown to treat EAE in mice ³² and prevent restenosis in rats ²² and angiogenesis in mice ²⁸ ; K _{Ca} 3.1 blockers reduce experimental brain oedema and attenuate traumatic brain injury ³³ ; K _{Ca} 3.1 openers are considered as potential therapeutics for cystic fibrosis and chronic obstructive pulmonary disease ¹¹
Comments	Voltage-independent calmodulin is also involved in trafficking ³⁴ ; intron-exon structure shared with $K_{Ca}2.1-K_{Ca}2.3$ (SK channels)

aa, amino acids; chr., chromosome; NS309, 6,7-dichloro-1H-indole-2,3-dione-3-oxime; IK, intermediate-conductance K+ channel; EBIO, 1-ethyl-2-benzimidazolinone; DCEBIO, 5,6-dichloro-1-ethyl-1,3-dihydro-2H-benzimidazol-2-one; ChTX, charybdotoxin; ShK, ShK toxin, a potassium channel blocker from the sea anemone Stichodactyla helianthus; BgK, BgK toxin, a potassium channel blocker from the sea anemone Bunodosoma granulifera; EAE, experimental autoimmune encephalomyelitis; ŠK, small-conductance K⁺ channel; ICA17043, bis(4-fluorophenyl)phenyl acetamide; UCL1608, 1-[(9-benzyl)fluoren-9-yl]-4-(hexahydro-1H-azepin-1-yl)but-2-yne hydrogen oxalate; IBMX, 3-isobutyl-1-methylxanthine; TEA, tetraethylammonium; EDHF, endothelium-derived hyperpolarizing factor.

1. Joiner WJ, Wang LY, Tang MD, and Kaczmarek LK (1997) hSK4, a member of a novel subfamily of calcium-activated potassium channels. Proc Natl Acad Sci USA 94:11013-11018

2. Ishii TM, Silvia C, Hirschberg B, Bond CT, Adelman JP, and Maylie J (1997) A human intermediate conductance calcium-activated potassium channel. Proc Natl Acad Sci USA 94:11651-11656.

3. Logsdon NJ, Kang J, Togo JA, Christian EP, and Aiyar J (1997) A novel gene, hKCa4, encodes the calcium-activated potassium channel in human T lymphocytes. J Biol Chem 272:32723-32726.

4. Ghanshani S, Wulff H, Miller MJ, Rohm H, Neben A, Gutman GA, Cahalan MD, and Chandy KG (2000) Up-regulation of the IKCa1 potassium channel during T-cell activation. Molecular mechanism and functional consequences. J Biol Chem 275:37137-37149.

5. Ghanshani S, Coleman M, Gustavsson P, Wu AC, Gargus JJ, Gutman GA, Dahl N, Mohrenweiser H, and Chandy KG (1998) Human calcium-activated potassium

channel gene KCNN4 maps to chromosome 19q13.2 in the region deleted in diamond-black fan anemia. *Genomics* 51:160–161. 6. Fanger CM, Ghanshani S, Logsdon NJ, Rauer H, Kalman K, Zhou J, Beckingham K, Chandy KG, Cahalan MD, and Aiyar J (1999) Calmodulin mediates calcium-dependent activation of the intermediate conductance KCa channel, IKCa1. *J Biol Chem* **274**:5746–5754.

7. Gardos G (1958) The function of calcium in the potassium permeability of human erythrocytes. Biochim Biophys Acta 30:653-654.

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9. Pena TL, Chen SH, Konieczny SF, and Rane SG (2000) Ras/MEK/ERK up-regulation of the fibroblast K_{Ca} channel FIK is a common mechanism for basic fibroblast growth factor and transforming growth factor-beta suppression of myogenesis. J Biol Chem 275:13677-13682.

10. Strobaek D, Teuber L, Jorgensen TD, Ahring PK, Kaer K, Hansen RS, Olesen SP, Christophersen P, and Skaaning-Jensen B (2004) Activation of human IK and SK Ca²⁺ -activated K⁺ channels by NS309 (6,7-dichloro-1H-indole-2,3-dione3-oxime). *Biochim Biophys Acta* **1665**:1–5.

11. Singh S, Syme CA, Singh AK, Devor DC, and Bridges RJ (2001) Benzimidazolone activators of chloride secretion: potential therapeutics for cystic fibrosis and chronic obstructive pulmonary disease. J Pharmacol Exp Ther 296:600-611. 12. Schroder RL, Jensen BS, Strobaek D, Olesen SP, and Christophersen P (2000) Activation of the human, intermediate-conductance, Ca²⁺-activated K⁺ channel by

- methylxanthines. Pflugers Arch Eur J Physiol 440:809-818.
- 13. Rauer H, Lanigan MD, Pennington MW, Aiyar J, Ghanshani S, Cahalan MD, Norton RS, and Chandy KG (2000) Structure-guided transformation of charybdotoxin yields an analog that selectively targets Ca²⁺-activated over voltage-gated K⁺ channels. J Biol Chem **275**:1201–1208. 14. Chandy KG, Wulff H, Beeton C, Pennington M, Gutman GA, and Cahalan MD (2004) K⁺ channels as targets for specific immunomodulation. Trends Pharmacol Sci **25**:280–289.
- 15. Wulff H, Miller MJ, Haensel W, Grissmer S, Cahalan MD, and Chandy KG (2000) Design of a potent and selective inhibitor of the intermediate-conductance Ca² ²⁺-activated K⁺ channel, IKCa1: a potential immunosuppressant. Proc Natl Acad Sci USA 97:8151-8156.
 16. Urbahns K, Horvath E, Stasch JP, and Mauler F (2003) 4-Phenyl-4H-pyrans as IK (Ca) channel blockers. Bioorg Med Chem Lett 13:2637-2639.
- 17. Stocker JW, De Franceschi L, McNaughton-Smith GA, Corrocher R, Beuzard Y, and Brugnara C (2003) ICA-17043, a novel Gardos channel blocker, prevents sickled red blood cell dehydration in vitro and in vivo in SAD mice. Blood 101:2412-2418.
- 18. Roxburgh CJ, Ganellin CR, Athmani S, Bisi A, Quaglia W, Benton DC, Shiner MA, Malik-Hall M, Haylett DG, and Jenkinson DH (2001) Synthesis and
- structure-activity relationships of cetiedil analogues as blockers of the Ca²⁺-activated K⁺ permeability of erythrocytes. *J Med Chem* 44:3244–3253. 19. Vandorpe DH, Shmukler BE, Jiang L, Lim B, Maylie J, Adelman JP, de Franceschi L, Cappellini MD, Brugnara C, and Alper SL (1998) cDNA cloning and functional characterization of the mouse Ca²⁺-gated K⁺ channel, mIK1Roles in regulatory volume decrease and erythroid differentiation. *J Biol Chem* **273**:21542–21553.
- 20. Ayabe T, Wulff H, Darmoul D, Cahalan MD, Chandy KG, and Ouellette AJ (2002) Modulation of mouse Paneth cell alpha-defensin secretion by mIKCa1, a ²⁺ activated, intermediate conductance potassium channel. J Biol Chem **277:**3793–3800. Ca24
- 21. Meyer R, Schonherr R, Gavrilova-Ruch O, Wohlrab W, and Heinemann SH (1999) Identification of ether a go-go and calcium-activated potassium channels in human melanoma cells. J Membr Biol 171:107-115.
- 22. Kohler R, Wulff H, Eichler I, Kneifel M, Neumann D, Knorr A, Grgic I, Kampfe D, Si H, Wibawa J, et al. (2003) Blockade of the intermediate-conductance calcium-activated potassium channel as a new therapeutic strategy for restenosis Circulation 108:1119-1125.
- 23. Eichler I, Wibawa J, Grgic I, Knorr A, Brakemeier S, Pries AR, Hoyer J, and Kohler R (2003) Selective blockade of endothelial Ca²⁺-activated small- and intermediate-conductance K⁺-channels suppresses EDHF-mediated vasodilation. Br J Pharmacol 138:594-601.
- 24. Brugnara C, Gee B, Armsby CC, Kurth S, Sakamoto M, Rifai N, Alper SL, and Platt OS (1996) Therapy with oral clotrimazole induces inhibition of the Gardos channel and reduction of erythrocyte dehydration in patients with sickle cell disease. J Clin Investig 97:1227-1234.
- 25. Jensen BS, Odum N, Jorgensen NK, Christophersen P, and Olesen SP (1999) Inhibition of T cell proliferation by selective block of Ca (2+)-activated K (+) channels. Proc Natl Acad Sci USA 96:10917-10921.
- 26. Khanna R, Chang MC, Joiner WJ, Kaczmarek LK, and Schlichter LC (1999) hSK4/hIK1, a calmodulin-binding K_{Ca} channel in human T lymphocytes. Roles in proliferation and volume regulation. J Biol Chem 274:14838-14849.
- 27. Kohler R, Degenhardt C, Kuhn M, Runkel N, Paul M, and Hoyer J (2000) Expression and function of endothelial Ca2+-activated K+ channels inhuman mesenteric artery: a single-cell reverse transcriptase-polymerase chain reaction and electrophysiological study in situ. Circ Res 87:496-503.
- 28. Grgic I, Eichler I, Heinau P, Si H, Brakemeier S, Hoyer J, and Kohler R (2005) Selective blockade of the intermediate-conductance Ca²⁺-activated K⁺ channel suppresses proliferation of microvascular and macrovascular endothelial cells and angiogenesis in vivo. Arterioscler Thromb Vasc Biol 25:704-709. 29. Begenisich T, Nakamoto T, Ovitt CE, Nehrke K, Brugnara C, Alper SL, and Melvin JE (2004) Physiological roles of the intermediate conductance, Ca²⁺-activatedpotassium channel Kcnn4. J Biol Chem **279:**47681–47687.
- 30. Rufo PA, Merlin D, Riegler M, Ferguson-Maltzman MH, Dickinson BL, Brugnara C, Alper SL, and Lencer WI (1997) The antifungal antibiotic, clotrimazole, inhibits
- chloride secretion by human intestinal T84 cells via blockade of distinct basolateral K⁺ conductances. Demonstration of efficacy in intact rabbit colon and in an in vivo mouse model of cholera. J Clin Investig 100:3111-3120.
- 31. Coghlan MJ, Carrol WA, and Gopalakrishnan M (2001) Recent developments in the biology and medicinal chemistry of potassium channel modulators: update from a decade of progress. J Med Chem 44:1-27. 32. Reich EP, Cui L, Yang L, Pugliese-Sivo C, Golovko A, Petro M, Vassileva G, Chu I, Nomeir AA, Zhang LK, et al. (2005) Blocking ion channel KCNN4 alleviates the
- symptoms of experimental autoimmune encephalomyelitis in mice. Eur J Immunol 35:1027-1036.
- 33. Mauler F, Hinz V, Horvath E, Schuhmacher J, Hofmann HA, Wirtz S, Hahn MG, and Urbahns K (2004) Selective intermediate/small-conductance calcium-activated potassium channel (KCNN4) blockers are potent and effective therapeutics in experimental brain oedema and traumatic brain injury caused by acute subdural haematoma. Eur J Neurosci 20:1761–1768
- 34. Joiner WJ, Khanna R, Schlichter LC, and Kaczmarek LK (2001) Calmodulin regulates assembly and trafficking of SK4/IK1 Ca²⁺-activated K⁺ channels. J Biol Chem 276:37980-37985.

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 $K_{Ca}4.1$ channels

Heteromeric association between rat Slack and Slo1,³ no β -subunits identified

Human: 1256aa NM_020822, chr. 9q34.3, KCNT1

Mouse: XM_622105 (predicted), chr. 2 Rat: 1237aa, NM_021853, chr. 3p13

Sodium-activated potassium channel, rat (Slack) ortholog gated by voltage and synergistically by

25-65pS (Slack),³ 60-180pS (Slack/Slo1 heteromeric channels)³; 88pS (80 mM symmetric K⁺),

165pS (160 mM symmetric K^+), prominent multiple subconductance states (*Slack*)⁶

Р <	Channel name Description
2	Other names Molecular information
	Associated subunits

Functional assays

Conductance Ion selectivity

Current

Ion selectivity	K -selective
Activation	Gated by voltage (weakly voltage-sensitive) and synergistically by internal Na^+ and CI^- (half-
	maximal Na ⁺ activation $[Na^+]_{0.5} = 15 \text{ mM}$ with 160 mM CI ⁻ ; half-maximal CI ⁻ activation
	$[Cl^{-}]_{0.5} = 8.1 \text{ mM with 80 mM Na}^{+})^{6}$
Inactivation	None
Activators	None
Blockers	TEA, $>60\%$ block by 20 mM ² ; quindine, $>90\%$ block by 1.0 mM ²
Gating inhibitors	Intracellular Ca ⁺² (5-fold reduction of NP ₀ increasing Ca ²⁺ from 0–3 μ M) ³
Radioligands	None
Channel distribution	Brain, testis, kidney (mouse Slo2.2) ⁶ ; brain [brainstem (red nucleus, oculomotor nucleus,
	mesencephalic trigeminal, trapezoid nucleus, gigantocellularius, vestibular nucleus), olfactory
	bulb, frontal cortex, hippocampus], kidney, testis (rat <i>Slack</i>) ¹ ; neuronal immunohistochemical
	staining observed in cell bodies and axonal tracts
Physiological functions	Not established
Mutations and pathophysiology	Not established; C. elegans slo-2 loss-of-function mutants hypersensitive to hypoxic death ^{$5,6$}
Pharmacological significance	Not established; native K_{Na} channels proposed to protect against hypoxic insult in cardiac muscles ⁴
Comments	No published functional expression data for the human ortholog

aa, amino acids; chr., chromosome; TEA, tetraethylammonium.

 $K_{Ca}4.1$

K⁺-selective

 K^+ -selective

internal Na⁺ and CI⁻ Slack, Slo2.2, KCNT1

Voltage-clamp, patch-clamp

 Bhattacharjee A, Gan L, and Kaczmarek LK (2002) Localization of the Slack potassium channel in the rat central nervous system. J Comp Neurol 454:241-254.
 Bhattacharjee A, Joiner WJ, Wu M, Ynag Y, Sigworth FJ, and Kaczmarek LK (2003) Slick (Slo2.1), a rapidly-gated sodium-activated potassium channel inhibited by ATP. J Neurosci 23:11681-11691.

3. Joiner WJ, Tang MD, Wang LY, Dworetzky SI, Boissard CG, Gan L, Gribkoff VK, and Kaczmarek LK (1998) Formation of intermediate-conductance-activated potassium channels by interaction of Slack and Slo subunits. *Nat Neurosci* 1:462-469.

4. Kameyama M, Kakei M, Sato R, Shibasaki T, Matsuda H, and Irisawa H (1984) Intracellular Na⁺ activates a K⁺ channel in mammalian cardiac cells. Nature (Lond)
309:354-356.
5. Yuan A, Dourado M, Butler A, Walton N, Wei A, and Salkoff L (2000) SLO-2, A K⁺ channel with an unusual Cl⁻ dependence. Nat Neurosci 3:771-779.

Yuan A, Dourado M, Butler A, Walton N, Wei A, and Salkoff L (2000) SLO-2, A K channel with an unusual CI dependence. Nat Neurosci 3:7/1-7/19.
 Yuan A, Santi CM, Wei A, Wang Z-W, Pollak K, Nonet M, Kaczmarek L, Crowder CM, and Salkoff L (2003) The sodium-activated potassium channel is encoded by a member of the Slo gene family. Neuron 37:765-773.

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TAI	BLE 8
$K_{Ca}4.2$	channels

1

	Γ_{Ca} : \mathcal{L} chambers
Channel Description Other names Molecular information	K _{Ca} 4.2 Sodium-activated potassium channel gated by voltage, internal Na ⁺ and Cl ⁻ , and inhibited by ATP Slick, Slo2.1, KCNT2 Human: 1138aa, NM_198503; chr. 1q31.3, KCNT2 Mouse: 1131aa, XM 136252, chr. 1
Associated subunits Functional Assays Current Conductance Ion Selectivity Activation	 Rat: 1142aa, NM_198762, chr. 13q13 No β-subunits identified; binding to PSD-95 scaffolding protein via first PDZ domain⁴ Voltage-clamp, patch-clamp K⁺-selective 141pS (130 mM symmetric K⁺), multiple subconductance states¹ K⁺-selective Gated by voltage (weakly voltage-sensitive) and synergistically by internal Na⁺ and Cl⁻ (5-fold increase in NP_o when Na⁺ raised from 1–100 mM, with 30 mM Cl⁻; 5-fold increase in NP_o when Cl⁻ raised from 3–130 mM, with 5 mM Na⁺)¹
Inactivation	None
Activators	None
Blockers	TEA, $>60\%$ block by 20 mM; quindine, $>90\%$ block by 1.0 mM ¹
Gating inhibitors	Intracellular ATP, >80% block by 5.0 mM
Radioligands	None
Channel distribution	Ubiquitous (mouse <i>Slo2.1</i>) ⁵ ; brain (olfactory bulb, supraoptic nucleus, hippocampus, somatosensory and visual cortex, thalamus, deep cerebellar nucleus, oculomotor nucleus, auditory nuclei), heart ² (rat <i>Slick</i>); neuronal immunohistochemical staining observed in cell bodies and axonal tracts
Physiological functions	Not established
Mutations and pathophysiology	Not established; C. elegans slo-2 loss-of-function mutants are hypersensitive to hypoxic death ^{4,5}
Pharmacological significance	Not established; native K_{Na} channels proposed to protect against hypoxic insult in cardiac muscles ³

aa, amino acids; chr., chromosome; PDZ, postsynaptic density 95/disc-large/zona occludens; TEA, tetraethylammonium.

1. Bhattacharjee A, Gan L, and Kaczmarek LK (2002) Localization of the Slack potassium channel in the rat central nervous system. J Comp Neurol 454:241-254. 2. Bhattacharjee A, Joiner WJ, Wu M, Yang Y, Sigworth FJ, and Kaczmarek LK (2003) Slick (Slo2.1), a rapidly-gated sodium-activated potassium channel inhibited by ATPJ. Neurosci 23:11681–11691.

3. Kameyama M, Kakei M, Sato R, Shibasaki T, Matsuda H, and Irisawa H (1984) Intracellular Na+ activates a K+ channel in mammalian cardiac cells. *Nature (Lond)* 309:354–356.

 Yuan A, Dourado M, Butler A, Walton N, Wei A, and Salkoff L (2000) SLO-2, a K⁺ channel with an unusual Cl⁻ dependence Nat Neurosci 3:771–779.
 Yuan A, Santi CM, Wei A, Wang Z-W, Pollak K, Nonet M, Kaczmarek L, Crowder CM, and Salkoff L (2003) The sodium-activated potassium channel is encoded by a member of the Slo gene family. Neuron 37:765-773.

TABLE 9

$K_{Ca}5.1$ channels			
Channel	K _{Ca} 5.1		
Description	pH-sensitive large-conductance potassium channel		
Other names	Slo3, KCNMC1, Kcnma3		
Molecular information	Human: BC028701 (coding sequence not defined), chr. 8p11.2, ⁷ KCNU1		
	Mouse: 1112aa, NM_008432, chr. 8		
	Rat: 1243aa, NM_031828, chr. 15p16		
Associated subunits	No β -subunits identified		
Functional Assays	Voltage- and patch-clamp		
Current	K^+ -selective (mouse $Slo3$)		
Conductance	106pS with 160 mM symmetric K^+ (mouse $Slo3$) ²		
Ion Selectivity	$P_{K+}/P_{Na+} = 5.0 \text{ (mouse } Slo3)^2$		
Activation	Gated by voltage and internal alkalization (half-maximal activation at pH 7.5) ¹⁻⁶		
Inactivation	None		
Activators	None		
Blockers	TEA, 50% block by 49 mM ²		
Gating inhibitors	None		
Radioligands	None		
Channel distribution	Testis, spermatocytes ²		
Physiological functions	Not established		
Mutations and pathophysiology	Not established		
Pharmacological significance	Not established		
Comments	No published functional expression data for the human ortholog		

aa, amino acids; chr., chromosome; TEA, tetraethylammonium.

1. Moss BL and Magleby KL (2001) Gating and conductance properties of BK channels are modulated by the S9-S10 tail domain of the alpha subunitA study of mSlo1 and mSlo3 wild-type and chimeric channels J Gen Physiol 118:711-734.

2. Schreiber M, Wei A, Yuan A, Gaut J, Saito M, and Salkoff L (1998) Slo3, a novel pH-sensitive K+ channel from mammalian spermatocytes. J Biol Chem 273:3509–3516. Schreiber M, Yuan A, and Salkoff L (1999) Transplantable sites confer calcium sensitivity to BK channels. Nat Neurosci 2:416-421.
 Shi J and Cui J (2001) Intracellular Mg²⁺ enhances the function of BK-type Ca²⁺-activated K⁺ channels. J Gen Physiol 118:589-606.
 Shi J, Krishnamoorthy G, Yang Y, Hu L, Chaturvedi N, Harilal D, Qin J, and Cui J (2002) Mechanism of magnesium activation of calcium-activated potassium channels.

Nature (Lond) 418:876-880.

6. Xia XM, Zhang X, and Lingle CJ (2004) Ligand-dependent activation of Slo family channels is defined by interchangeable cytosolic domains. J Neurosci 24:5585-5591. 7. GenBank accession no. AP000074.



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